# Package: aIc (via r-universe)

September 14, 2024

**Title** Testing for Compositional Pathologies in Datasets Version 1.0 Date 2022-09-27 Author Greg Gloor Maintainer Greg Gloor <ggloor@uwo.ca> **Description** A set of tests for compositional pathologies. Tests for coherence of correlations with aIc.coherent() as suggested by (Erb et al. (2020) <doi:10.1016/j.acags.2020.100026>), compositional dominance of distance with aIc.dominant(), compositional perturbation invariance with aIc.perturb() as suggested by (Aitchison (1992) <doi:10.1007/BF00891269>) and singularity of the covariation matrix with aIc.singular(). Currently tests five data transformations: prop, clr, TMM, TMMwsp, and RLE from the R packages 'ALDEx2', 'edgeR' and 'DESeq2' (Fernandes et al (2014) < doi:10.1186/2049-2618-2-15>, Anders et al. (2013)<doi:10.1038/nprot.2013.099>). License GPL (>= 3)**Encoding** UTF-8 URL https://github.com/ggloor/alc BugReports https://github.com/ggloor/aIc/issues RoxygenNote 7.2.1 VignetteBuilder knitr Imports matrixcalc, zCompositions, shiny, edgeR, ALDEx2, vegan Suggests BiocStyle, knitr, rmarkdown **Depends** R (>= 3.5.0), **Repository** https://ggloor.r-universe.dev RemoteUrl https://github.com/ggloor/aic RemoteRef HEAD

**RemoteSha** ada8f7e396ebb275dcd22b2af6123681dd1c8b4e

Type Package

2 alc.coherent

## **Contents**

aIc.coherent			Ca a g				ıpa	os	iti	on	al	co	he	ere	no	ce	of	SC	ım	pl	les	i ii	n c	а с	lai	ta.	sei	t fe	or
Index																													12
	transcriptome .				 •	•	 ٠	•					•					•	•		•				•				11
	singleCell																												
	selex																												
	metaTscome																												
	alc.singular meta16S																												
	aIc.scale																												
	aIc.runExample																												
	aIc.plot																												6
	aIc.perturb																												
	alc.dominant .																												
	alc.coherent				 																								2

## Description

'alc.coherent' compares the correlation coefficients of features in common of the full dataset and a subset of the dataset. This is expected to be false for all compositional datasets and transforms.

## Usage

```
alc.coherent(
  data,
  norm.method = "prop",
  zero.remove = 0.95,
  zero.method = "prior",
  log = FALSE,
  group = NULL,
  cor.test = "spearman"
)
```

## Arguments

data	can be any dataframe or matrix with samples by column
norm.method	can be prop, clr, RLE, TMM, TMMwsp, lvha, iqlr
zero.remove	is a value. Filter data to remove features that are 0 across at least that proportion of samples: default $0.95$
zero.method	can be any of NULL, prior, GBM or CZM. NULL will not impute or change 0 values, GBM and CZM are from the zCompositions R package, and prior will simply add 0.5 to all counts.

aIc.dominant 3

log	is a logical. log transform the prop, RLE or TMM outputs, default=FALSE
group	is a vector containing group information. Required for clr, RLE,
cor.test	is either the pearson or spearman method (default)

#### Value

Returns a list with the correlation in cor, a yes/no binary decision in is.coherent, the x and y values for a scatterplot of the correlations in the full and subcompositions, and the plot and axis labels in main xlab and ylab.

### Author(s)

Greg Gloor

## **Examples**

```
data(selex)
group = c(rep('N', 7), rep('S', 7))
x <- aIc.coherent(selex, group=group, norm.method='clr', zero.method='prior')
plot(x$plot[,1], x$plot[,2], main=x$main, ylab=x$ylab, xlab=x$xlab)</pre>
```

aIc.dominant

alc.dominant calculates the subcompositional dominance of a sample in a dataset for a given correction. This compares the distances of samples of the full dataset and a subset of the dataset. This is expected to be true if the transform is behaving rationally in compositional datasets.

## **Description**

alc.dominant calculates the subcompositional dominance of a sample in a dataset for a given correction. This compares the distances of samples of the full dataset and a subset of the dataset. This is expected to be true if the transform is behaving rationally in compositional datasets.

## Usage

```
alc.dominant(
  data,
  norm.method = "prop",
  zero.remove = 0.95,
  zero.method = "prior",
  log = FALSE,
  distance = "euclidian",
  group = NULL
)
```

4 aIc.perturb

## **Arguments**

data	can be any dataframe or matrix with samples by column
norm.method	can be prop, clr, RLE, TMM, TMMwsp
zero.remove	is a value. Filter data to remove features that are 0 across at least that proportion of samples: default 0.95
zero.method	can be any of NULL, prior, GBM or CZM. NULL will not impute or change 0 values, GBM (preferred) and CZM are from the zCompositions R package, and prior will simply add 0.5 to all counts.
log	is a logical. log transform the RLE or TMM outputs, default=FALSE
distance	can be euclidian, bray, or jaccard. euclidian on log-ratio transformed data is the same as the Aitchison distance. default=euclidian
group	is a vector containing group information. Required for clr, RLE, TMM, lvha,

and iqlr based normalizations.

datasets.

#### Value

Returns a list with the overlap between distances in the full and subcomposition in ol (expect 0), a yes/no binary decision in is.dominant and the table of distances for the whole and subcomposition in dist.all and dist.sub, a plot showing a histogram of the resulting overlap in distances in plot, and the plot and axis labels in main xlab and ylab

#### Author(s)

Greg Gloor

## **Examples**

### **Description**

aIc.perturb calculates the perturbation invariance of distance for samples with a given correction. This compares the distances of samples of the full dataset and a the perturbed dataset. This is expected to be true if the transform is behaving rationally in compositional datasets.

aIc.perturb 5

## Usage

```
alc.perturb(
  data,
  norm.method = "prop",
  zero.remove = 0.95,
  zero.method = "prior",
  distance = "euclidian",
  log = FALSE,
  group = NULL
)
```

#### **Arguments**

data can be any dataframe or matrix with samples by column

norm.method can be prop, clr, RLE, TMM, TMMwsp

zero.remove is a value. Filter data to remove features that are 0 across at least that proportion

of samples: default 0.95

zero.method can be any of NULL, prior, GBM or CZM. NULL will not impute or change 0

values, GBM (preferred) and CZM are from the zCompositions R package, and

prior will simply add 0.5 to all counts.

distance can be euclidian, bray, or jaccard. euclidian on log-ratio transformed data is the

same as the Aitchison distance. default=euclidian

log is a logical. log transform the RLE or TMM outputs, default=FALSE

group is a vector containing group information. Required for clr, RLE, TMM, lvha,

and iqlr based normalizations.

## Value

Returns a list with the maximum proportional perturbation in ol (expect 0, but values up to 1 is.perturb, the table of distances for the whole and perturbation in dist.all and dist.perturb, the histogram of the perturbations in plot, and the plot and axis labels in main xlab and ylab.

#### Author(s)

Greg Gloor

#### **Examples**

```
data(selex) group = c(rep('N', 7), rep('S', 7)) x <- aIc.perturb(selex, group=group, norm.method='clr', distance='euclidian', zero.method='prior') plot(x plot, main=x main, ylab=x ylab, xlab=x xlab)
```

6 alc.runExample

aIc.plot

alc.plot plots the result of the distance tests.

## **Description**

aIc.plot plots the result of the distance tests.

## Usage

```
aIc.plot(test.out)
```

## **Arguments**

test.out

is the output from either aIc.dominant, aIc.scale, aIc.perturb

## Value

returns a plot of the density of the distance test results. test result.

#### Author(s)

Greg Gloor

## **Examples**

```
data(selex)
group = c(rep('N', 7), rep('S', 7))
test.out <- aIc.dominant(selex, norm.method='prop', group=group)
aIc.plot(test.out)</pre>
```

aIc.runExample

alc.runExample loads the associated shiny app This will load the selex example dataset with the default group sizes, the user can upload their own local dataset and adjust groups accordingly.

## **Description**

alc.runExample loads the associated shiny app This will load the selex example dataset with the default group sizes, the user can upload their own local dataset and adjust groups accordingly.

## Usage

```
aIc.runExample()
```

aIc.scale 7

#### Value

No return value, but instead opens a shiny connection to your default web browser with the selex dataset as an example.

#### Author(s)

Greg Gloor

## **Examples**

```
library(aIc)
aIc.runExample()
```

aIc.scale

alc.scale calculates the scaling invariance of a sample in a dataset for a given correction. This compares the distances of samples of the full dataset and a scaled version of the dataset. This is expected to be true if the transform is behaving rationally in compositional datasets.

#### **Description**

aIc.scale calculates the scaling invariance of a sample in a dataset for a given correction. This compares the distances of samples of the full dataset and a scaled version of the dataset. This is expected to be true if the transform is behaving rationally in compositional datasets.

### Usage

```
alc.scale(
  data,
  norm.method = "prop",
  zero.remove = 0.95,
  zero.method = "prior",
  distance = "euclidian",
  log = FALSE,
  group = NULL
)
```

## **Arguments**

data can be any dataframe or matrix with samples by column

norm.method can be prop, clr, iqlr, lvha, RLE, TMM, TMMwsp

zero.remove is a value. Filter data to remove features that are 0 across at least that proportion of samples: default 0.95

zero.method can be any of NULL, prior, GBM or CZM. NULL will not impute or change 0 values, GBM (preferred) and CZM are from the zCompositions R package, and prior will simply add 0.5 to all counts.

8 alc.singular

distance	can be euclidian, bray, or jaccard. euclidian on log-ratio transformed data is the same as the Aitchison distance. default=euclidian
log	is a logical. log transform the RLE or TMM outputs, default=FALSE
group	is a vector containing group information. Required for clr, RLE, TMM, lvha, and iqlr based normalizations.

#### Value

Returns a list with the overlap between distances in the full and scaled composition in ol (expect 0), a yes/no binary decision in is.scale and the table of distances for the whole and scaled composition in dist.all and dist.scale, a plot showing a histogram of the resulting overlap in distances in plot, and the plot and axis labels in main xlab and ylab

#### Author(s)

Greg Gloor

## **Examples**

```
data(selex)
group = c(rep('N', 7), rep('S', 7))
x <- aIc.scale(selex, group=group, norm.method='clr', zero.method='prior')
plot(x$plot, main=x$main, ylab=x$ylab, xlab=x$xlab)</pre>
```

aIc.singular

alc.singular tests for singular data. This is expected to be true if the transform is behaving rationally in compositional datasets and also true in the case of datasets with more features than samples.

#### **Description**

alc.singular tests for singular data. This is expected to be true if the transform is behaving rationally in compositional datasets and also true in the case of datasets with more features than samples.

## Usage

```
alc.singular(
  data,
  norm.method = "prop",
  zero.remove = 0.95,
  zero.method = "prior",
  log = FALSE,
  group = NULL
)
```

meta16S 9

## **Arguments**

data can be any dataframe or matrix with samples by column

norm.method can be prop, clr, RLE, TMM, TMMwsp

zero.remove is a value. Filter data to remove features that are 0 across at least that proportion

of samples: default 0.95

zero.method can be any of NULL, prior, GBM or CZM. NULL will not impute or change 0

values, GBM (preferred) and CZM are from the zCompositions R package, and

prior will simply add 0.5 to all counts.

log is a logical. log transform the RLE or TMM outputs, default=FALSE

group is a vector containing group information. Required for clr, RLE, TMM, lvha,

and iqlr based normalizations.

#### Value

Returns a list with a yes/no binary decision in is.singular and the covariance matrix in cov.matrix

#### Author(s)

Greg Gloor

## **Examples**

```
data(selex)
group = c(rep('N', 7), rep('S', 7))
x <- aIc.singular(selex, group=group, norm.method='clr', zero.method='prior')</pre>
```

meta16S

16S rRNA tag-sequencing data

## **Description**

A count table of a 16S rRNA amplicon data Two groups, pupils and centenarians are represented with 198 and 161 samples per group respectively. samples are by column and OTU ids are by row.

## Usage

```
data(meta16S)
```

#### **Format**

A data frame with 359 columns and 860 rows

#### Source

doi: 10.1128/mSphere.00327-17

10 selex

metaTscome

meta-transcriptome data

## **Description**

A count table of a mixed population or metatranscriptome experiment. Two groups, H and BV are represented with 7 and 10 samples per group respectively. samples are by column and functions are by row.

## Usage

```
data(metaTscome)
```

#### **Format**

A data frame with 17 columns and 3647 rows

#### **Source**

doi:10.1007/978-3-030-71175-7\_17 and doi:10.1007/978-1-4939-8728-3\_13

selex

Selection-based differential sequence variant abundance dataset

## Description

This data set gives the differential abundance of 1600 enzyme variants grown under selective (NS) and selective (S) conditions

## Usage

```
data(selex)
```

## **Format**

A data frame with 14 columns and 1600 rows

### **Source**

DOI:10.1073/pnas.1322352111

singleCell 11

singleCell

single cell transcriptome data

## **Description**

A count table of a single cell transcriptome data subset from the count table from doi:10.1038/s41592-019-0372-4. Two groups memory T cells, and cytotoxic T cells, 1000 cells per group. samples are by column and genes are by row.

## Usage

```
data(singleCell)
```

#### **Format**

A data frame with 2000 columns and 1508 rows

#### **Source**

https://www.nature.com/articles/s41592-019-0372-4

transcriptome

Saccharomyces cerevisiae transcriptome

## **Description**

A count table of a highly replicated RNA-seq experiment with samples by column and genes by row. Two groups composed of SNF2 knockout and WT, 48 samples in each.

## Usage

```
data(transcriptome)
```

#### **Format**

A data frame with 96 columns and 5892 rows

#### Source

DOI: 10.1261/rna.053959.115 and PRJEB5348

# **Index**

```
\ast datasets
     meta16S, 9
     {\tt metaTscome},\, {\color{red}10}
     selex, 10
     singleCell, 11
     transcriptome, 11
aIc.coherent, 2
aIc.dominant, 3
aIc.perturb, 4
aIc.plot, 6
aIc.runExample, 6
aIc.scale, 7
\verb"aIc.singular", 8
meta16S, 9
metaTscome, 10
selex, 10
{\tt singleCell}, {\tt 11}
transcriptome, \\ 11
```